

Mouse IL-1 α ELISA Kit

EMIL1A EMIL1A2

1409.2

Number	Description
EMIL1A	<p>Mouse Interleukin-1 alpha (IL-1α) ELISA Kit, sufficient reagents for 96 determinations</p> <p>Kit Contents:</p> <p>Anti-Mouse IL-1α Precoated 96-well Strip Plate, 1 each</p> <p>Lyophilized Recombinant Mouse IL-1α Standard, 2 vials</p> <p>Standard Diluent, 12 ml, contains 0.1% sodium azide</p> <p>Biotinylated Anti-Mouse IL-1α Antibody Reagent, 8 ml, contains 0.1% sodium azide</p> <p>30X Wash Buffer, 50 ml</p> <p>Streptavidin-HRP Concentrate, 75 μl</p> <p>Streptavidin-HRP Dilution Buffer, 14 ml</p> <p>TMB Substrate Solution, 13 ml</p> <p>Stop Solution, 13 ml, contains 0.16 M sulfuric acid</p> <p>Adhesive Plate Covers, 6 each</p>
EMIL1A2	<p>Mouse Interleukin-1 alpha (IL-1α) ELISA Kit, sufficient reagents for 2 \times 96 determinations</p> <p>Kit Contents:</p> <p>Anti-Mouse IL-1α Precoated 96-well Strip Plate, 2 each</p> <p>Lyophilized Recombinant Mouse IL-1α Standard, 4 vials</p> <p>Standard Diluent, 2 \times 12 ml, contains 0.1% sodium azide</p> <p>Biotinylated Anti-Mouse IL-1α Antibody Reagent, 2 \times 8 ml, contains 0.1% sodium azide</p> <p>30X Wash Buffer, 2 \times 50 ml</p> <p>Streptavidin-HRP Concentrate, 2 \times 75 μl</p> <p>Streptavidin-HRP Dilution Buffer, 2 \times 14 ml</p> <p>TMB Substrate Solution, 2 \times 13 ml</p> <p>Stop Solution, 2 \times 13 ml, contains 0.16 M sulfuric acid</p> <p>Adhesive Plate Covers, 12 each</p>

For research use only. Not for use in diagnostic procedures.

Storage: For maximum stability, store in a non-defrosting -20°C freezer and refer to the expiration date for frozen storage on the label. Alternatively, store at 2-8°C and refer to the expiration date for refrigerated storage. Once thawed, store at 4°C until the expiration date for refrigerated storage. Kit is shipped on dry ice.




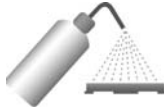


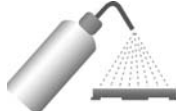




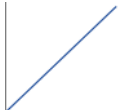
Introduction

The Thermo Scientific Mouse Interleukin-1 alpha (IL-1 α) ELISA is an *in vitro* enzyme-linked immunosorbent assay for the quantitative measurement of mouse IL-1 α in culture supernatants; EDTA, heparin and sodium citrate plasma; and serum.

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Procedure Summary

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1. Add 50 μ l of Biotinylated Antibody Reagent to each well. | 
2. Add 50 μ l of standards or samples to each well in duplicate. | 
3. Cover plate and incubate at room temperature (20-25°C) for 2 hours. | 
4. Wash plate THREE times. |
| 
5. Add 100 μ l of prepared Streptavidin-HRP Solution to each well. | 
6. Cover plate and incubate at room temperature for 30 minutes. | 
7. Wash plate THREE times. | 
8. Add 100 μ l of TMB Substrate Solution to each well. |
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9. Develop plate in the dark at room temperature for 30 minutes. | 
10. Stop reaction by adding 100 μ l of Stop Solution to each well. | 
11. Measure absorbance on a plate reader at 450 nm minus 550 nm. | 
12. Calculate results using graph paper or curve-fitting statistical software. |

Additional Materials Required

- Precision pipettors with disposable plastic tips to deliver 5-1,000 μ l
- Plastic pipettes to deliver 5-15 ml
- Ultrapure water
- A glass or plastic two-liter container to prepare Wash Buffer
- A squirt wash bottle or an automated 96-well plate washer
- 1.5 ml polypropylene or polyethylene tubes to prepare standards – do not use polystyrene, polycarbonate or glass tubes
- Disposable reagent reservoirs
- 15 ml plastic tube to prepare Streptavidin-HRP Solution
- Microcentrifuge to prepare Streptavidin-HRP Solution
- A standard ELISA reader for measuring absorbance at 450 nm and 550 nm. If a 550 nm filter is not available, the absorbance can be measured at 450 nm only. Refer to the instruction manual supplied with the instrument being used.
- Graph paper or a computerized curve-fitting statistical software package

Precautions

- **All samples and reagents must be at room temperature (20-25°C) before use in the ELISA.**
- Review all instructions carefully and verify all components against the Kit Contents list (page 1) before beginning.
- Do not use water baths to thaw samples. Thaw at room temperature.
- When assaying culture medium, prepare the standard curve and sample dilutions using the same medium used to culture the cells. For example, if RPMI with 10% fetal calf serum (FCS) was used to culture cells, then use RPMI with 10% FCS to dilute the standard and samples. For best results, use a culture medium that contains a carrier protein such as FCS. The lack of a carrier protein may affect the stability of the standards and compromise the standard curve. If serum-free media must be used, prepare two standard curves: one with carrier and one without. Compare curves to determine if lack of carrier caused degradation that resulted in a signal decrease. If the standard curve without carrier has an equivalent or slightly higher signal than the curve with the carrier, then serum-free media may be used. Alternatively, add carrier protein to both standards and samples.
- If using a multichannel pipettor, always use a new disposable reagent reservoir.
- Use new disposable pipette tips for each transfer to avoid cross-contamination.
- Use a new adhesive plate cover for each incubation step.
- Once reagents have been added to the plate, take care NOT to let plate DRY at any time during the assay.
- Avoid microbial contamination of reagents.
- Vigorous plate washing is essential.
- Avoid exposing reagents to excessive heat or light during storage and incubation.
- Do not mix reagents from different kit lots. Discard unused kit components.
- Do not use glass pipettes to measure TMB Substrate Solution. Take care not to contaminate the solution. If the solution is blue before use, DO NOT USE IT.
- Individual components may contain antibiotics and preservatives. Wear gloves while performing the assay to avoid contact with samples and reagents. Please follow proper disposal procedures.
- Some components of this kit contain sodium azide. Please dispose of reagents according to local regulations.

Sample Preparation

Sample Handling

- Serum; EDTA, heparin or sodium citrate plasma; or culture supernatants may be tested in this ELISA.
- 50 µl per well or serum, plasma or culture supernatant are required.
- Store samples to be assayed within 24 hours at 2-8°C. For long-term storage, aliquot and freeze samples at -70°C.
- Avoid repeated freeze-thaw cycles when storing samples.
- Test samples and standards must be assayed in duplicate each time the ELISA is performed.
- Gradually equilibrate samples to room temperature before beginning assay. Do not use heated water baths to thaw or warm samples.
- Mix samples by gently inverting tubes.
- If samples are clotted, grossly hemolyzed, lipemic or microbially contaminated, or if there is any question about the integrity of a sample, make a note on the template and interpret results with caution.

Sample Dilution

- If the mouse IL-1 α concentration possibly exceeds the highest point of the standard curve (i.e., 1,000 pg/ml), prepare one or more 10-fold dilutions of the test sample. When testing **culture supernatants**, prepare the serial dilutions using the culture medium (see Precautions section). When testing **serum or plasma**, prepare the serial dilutions using the Standard Diluent provided. For example, a 10-fold dilution is prepared by adding 50 µl of test sample to 450 µl of appropriate diluent. Mix thoroughly between dilutions before assaying.

Reagent Preparation

For procedural differences when using partial plates, look for **(PP)** throughout this instruction booklet.

Wash Buffer

1. Label a clean glass or plastic two-liter container "Wash Buffer." The 30X Wash Buffer may have a cloudy appearance.
2. Add the entire contents of one 30X Wash Buffer bottle (50 ml) to the two-liter container and dilute to a final volume of 1.5 liters with ultrapure water. Mix thoroughly.

(PP) When using partial plates, store the reconstituted Wash Buffer at 2-8°C.

Note: Wash Buffer must be at room temperature before use in the assay. Do not use Wash Buffer if it becomes visibly contaminated during storage.

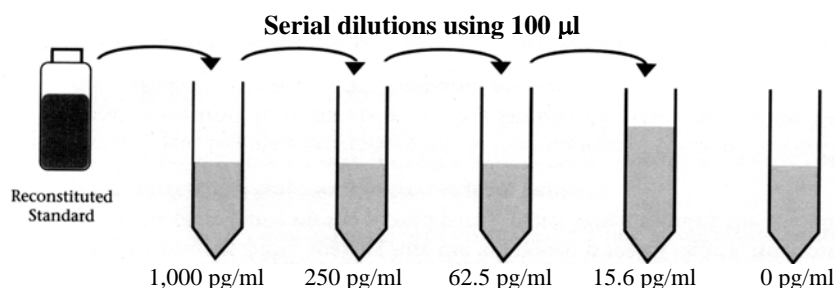
Standards

- **(PP)** Reconstitute and use one vial of the lyophilized standard per partial plate.
 - Prepare standards just before use and use within one hour of reconstitution. Do not store reconstituted standards.
1. When testing **culture supernatant samples**, reconstitute standard with ultrapure water. Reconstitution volume is stated on the standard vial label. The standard will dissolve in approximately 1 minute. Mix by gently inverting vial. Use the sample culture medium to prepare standard curve serial dilutions (see Precautions section).

When testing **serum or plasma samples**, reconstitute standard with ultrapure water. Reconstitution volume is stated on the standard vial label. The standard will dissolve in approximately 1 minute. Mix by gently inverting vial. Use the Standard Diluent provided to prepare standard curve serial dilutions.

Testing of RPMI with different concentrations of fetal bovine serum has shown that this ELISA is not adversely affected by culture medium. Therefore, when testing **serum or plasma and cell culture supernatants on the same plate**, reconstitute the standard with ultrapure water and make the 1:4 dilutions with Standard Diluent. If an unusual type of culture medium is being used, validate the medium to establish if the same standard curve can be used for the different sample types. Prepare a standard curve (including a zero/blank) using culture medium to reconstitute and dilute the standard. Use medium containing serum or other protein supplement to maximize stability of the mouse IL-1 α . Perform this curve in parallel with a standard curve reconstituted in ultrapure water and diluted in the Standard Diluent provided. If the OD values of the two curves are within 10% of the mean for both curves, then the assay can be performed with Standard Diluent, whether you are testing culture supernatant or serum samples.

2. Label five tubes, one for each standard curve point: 1,000, 250, 62.5, 15.6, and 0 pg/ml, then prepare 1:4 serial dilutions for the standard curve as follows:
3. Pipette 300 μ l of appropriate diluent into each tube.
4. Pipette 100 μ l of the reconstituted standard into the first tube (i.e., 1,000 pg/ml) and mix.
5. Pipette 100 μ l of this dilution into the second tube (i.e., 250 pg/ml) and mix.
6. Repeat the serial dilutions (using 100 μ l) two more times to complete the standard curve points. These concentrations, 1,000 pg/ml, 250 pg/ml, 62.5 pg/ml, 15.6 pg/ml, and 0 pg/ml, are the standard curve.



Assay Procedure

A. Biotinylated Antibody Reagent and Sample Incubation

- **(PP)** Determine the number of strips required. Leave these strips in the plate frame. Tightly seal remaining unused strips in the provided foil pouch with desiccant and store at 2-8°C. After completing assay, retain plate frame for second partial plate. When using second partial plate, place strips securely in the plate frame.
 - Use the Data Template provided to record the locations of the zero standard (blank or negative control), mouse IL-1 α standards and test samples. Perform five standard points and one blank in duplicate with each series of unknown samples.
 - If using a multichannel pipettor, use a new reagent reservoir to add the Biotinylated Antibody Reagent. Remove from the vial only the amount required for the number of strips being used.
1. Add 50 μ l of Biotinylated Antibody Reagent to each well.
 2. Add 50 μ l of reconstituted standards or test samples in duplicate to each well. Mix well by gently tapping the plate several times.

Note: If the mouse IL-1 α concentration in any test sample possibly exceeds the highest point on the standard curve, 1,000 pg/ml, see Sample Preparation – Sample Dilution section.

3. Add 50 μ l of Standard Diluent to all wells that do not contain standards or samples.
4. Carefully cover plate with an adhesive plate cover. Ensure all edges and strips are sealed tightly by running your thumb over edges and down each strip. Incubate for two (2) hours at room temperature, 20-25°C.
5. Carefully remove adhesive plate cover and wash **THREE** times with Wash Buffer as described in the Plate Washing section (section B).

B. Plate Washing

1. Gently squeeze the long sides of plate frame before washing to ensure all strips securely remain in the frame.
2. Discard plate contents. Use a squirt wash bottle to vigorously fill each well completely with Wash Buffer, then discard plate contents. Repeat procedure two additional times for a total of **THREE** washes. Blot plate onto paper towels or other absorbent material.

Note: For automated washing, aspirate all wells and wash **THREE** times with Wash Buffer, overfilling wells with Wash Buffer. Blot plate onto paper towels or other absorbent material.

C. Streptavidin-HRP Solution Preparation and Incubation

- Prepare Streptavidin-HRP Solution **immediately before use**. Do not prepare more solution than required.
 - Do not store prepared Streptavidin-HRP Solution.
 - Use a 15 ml plastic tube to prepare Streptavidin-HRP Solution.
 - If using a multichannel pipettor, **use new reagent reservoir and pipette tips** when adding the prepared Streptavidin-HRP Solution.
1. Briefly centrifuge the Streptavidin-HRP Concentrate to force entire vial contents to the bottom.
 2. **(PP)** Use only the Streptavidin-HRP Solution amount required for the number of strips being used. For each strip, mix 2.5 μ l of Streptavidin-HRP Concentrate with 1 ml of Streptavidin-HRP Dilution Buffer. Store Streptavidin-HRP Concentrate reserved for additional strips at 2-8°C.

For one complete 96-well plate, add 30 μ l of Streptavidin-HRP Concentrate to 12 ml of Streptavidin-HRP Dilution Buffer and mix gently.

3. Add 100 μ l of prepared Streptavidin-HRP Solution to each well.
4. Carefully attach a new adhesive plate cover, ensuring all edges and strips are tightly sealed. Incubate plate for 30 minutes at room temperature, 20-25°C.
5. Carefully remove the adhesive plate cover and wash **THREE** times as described in the Plate Washing section.

D. Substrate Incubation and Stop Step

- Use new disposable reagent reservoirs when adding TMB Substrate Solution and Stop Solution.
 - Dispense from bottle ONLY amount required, 100 µl per well, for the number of wells being used. Do not use a glass pipette to measure the TMB Substrate Solution.
 - **(PP)** Do not combine leftover substrate with that reserved for the second partial plate. Take care not to contaminate remaining TMB Substrate Solution.
1. Pipette 100 µl of TMB Substrate Solution into each well.
 2. Allow color reaction to develop at room temperature in the dark for 30 minutes. Do not cover plate with aluminum foil or a plate sealer. The substrate reaction yields a blue solution that turns yellow when Stop Solution is added.
 3. After 30 minutes, stop the reaction by adding 100 µl of Stop Solution to each well.

E. Absorbance Measurement

Note: Evaluate the plate within 30 minutes of stopping the reaction.

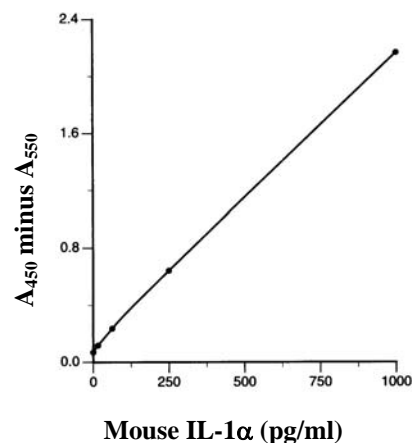
Measure the absorbance on an ELISA plate reader set at 450 nm and 550 nm. Subtract 550 nm values from 450 nm values to correct for optical imperfections in the microplate. If an absorbance at 550 nm is not available, measure the absorbance at 450 nm only.

Note: When the 550 nm measurement is omitted, absorbance values will be higher.

F. Calculation of Results

- The standard curve is used to determine mouse IL-1 α amount in an unknown sample. Generate the standard curve by plotting the average absorbance obtained for each standard concentration on the vertical (Y) axis vs. the corresponding mouse IL-1 α concentration (pg/ml) on the horizontal (X) axis.
- Calculate results using graph paper or curve-fitting statistical software. The mouse IL-1 α amount in each sample is determined by interpolating from the absorbance value (Y axis) to mouse IL-1 α concentration (X axis) using the standard curve.
- If the test sample was diluted, multiply the interpolated value obtained from the standard curve by the dilution factor to calculate pg/ml of mouse IL-1 α in the sample.
- Absorbance values obtained for duplicates should be within 10% of the mean value. Duplicate values that differ from the mean by greater than 10% should be considered suspect and repeated.

Example Standard Curve



Performance Characteristics

Sensitivity: <6 pg/ml

The sensitivity or Lower Limit of Detection (LLD)¹ is determined by assaying replicates of zero and the standard curve. The mean signal of zero + 2 standard deviations read in dose from the standard curve is the LLD. This value is the smallest dose that is not zero with 95% confidence.

Assay Range: 15.6-1,000 pg/ml

Suggested standard curve points are 1,000, 250, 62.5, 15.6, and 0 pg/ml.

Reproducibility:

Intra-assay CV: <10%

Inter-assay CV: <10%

Specificity: This ELISA is specific for the measurement of natural and recombinant mouse IL-1 α . This ELISA does not cross-react with human IL-1 α , human IL-1 β , or mouse IL-1 β , IL-3, IL-4, IL-5, IL-7, IL-10, TNF α , IFN γ , or GM-CSF.

Normal Levels: The average level of mouse-IL-1 α in 23 normal serum samples is 23 pg/ml with a range of 0 to 89 pg/ml. The average level of mouse IL-1 α in 12 normal plasma samples is 7.5 pg/ml with a range of 0 to 15 pg/ml.

Reference

1. *Immunoassay: A Practical Guide*, Chan and Perlstein, Eds., 1987, Academic Press: New York, p71.

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Data Templates

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
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H												

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