

Human IL-1 α ELISA Kit

EH2IL1A EH2IL1A2 EH2IL1A5

1375.3

Number	Description
EH2IL1A	Human Interleukin-1 alpha (IL-1α) ELISA Kit , sufficient reagents for 96 determinations Kit Contents: Anti-Human IL-1α Precoated 96-well Strip Plate , 1 each Lyophilized Recombinant Human IL-1α Standard , 2 vials Standard Diluent, 12 ml , contains 0.01% thimerosal Biotinylated Antibody Reagent , 8 ml, contains 0.01% thimerosal 30X Wash Buffer , 50 ml Streptavidin-HRP Concentrate , 75 μ l Streptavidin-HRP Dilution Buffer , 14 ml, contains 0.01% thimerosal TMB Substrate , 13 ml Stop Solution , 13 ml, contains 0.16 M sulfuric acid Adhesive Plate Covers , 6 each
EH2IL1A2	Human IL-1α ELISA Kit , sufficient reagents for 2 \times 96 determinations Kit Contents: Anti-Human IL-1α Precoated 96-well Strip Plate , 2 each Lyophilized Recombinant Human IL-1α Standard , 4 vials Standard Diluent , 2 \times 12 ml, contains 0.01% thimerosal Biotinylated Antibody Reagent , 2 \times 8 ml, contains 0.01% thimerosal 30X Wash Buffer , 2 \times 50 ml Streptavidin-HRP Concentrate , 2 \times 75 μ l Streptavidin-HRP Dilution Buffer , 2 \times 14 ml, contains 0.01% thimerosal TMB Substrate , 2 \times 13 ml Stop Solution , 2 \times 13 ml, contains 0.16 M sulfuric acid Adhesive Plate Covers , 12 each
EH2IL1A5	Human IL-1α ELISA Kit , sufficient reagents for 5 \times 96 determinations Kit Contents: Anti-Human IL-1α Precoated 96-well Strip Plate , 5 each Lyophilized Recombinant Human IL-1α Standard , 5 vials Standard Diluent , 75 ml Biotinylated Antibody Reagent , 35 ml 30X Wash Buffer , 200 ml Streptavidin-HRP Concentrate , 250 μ l Streptavidin-HRP Dilution Buffer , 70 ml TMB Substrate , 5 \times 13 ml Stop Solution , 55 ml, contains 0.16 M sulfuric acid Adhesive Plate Covers , 30 each

For research use only – not for use in diagnostic procedures.

Storage: For maximum stability, store in a non-defrosting -20°C freezer and refer to the expiration date for frozen storage on the label. Alternatively, store at 2-8°C and refer to the expiration date for refrigerated storage. Once thawed, store at 4°C until the expiration date for refrigerated storage. Kit is shipped on dry ice.





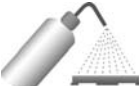


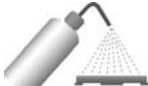




Introduction

This Thermo Scientific ELISA Kit is for quantification of human IL-1 α in culture supernatants, plasma (EDTA, heparin and sodium citrate), serum and urine.

Table of Contents

Introduction	2
Procedure Summary.....	2
Additional Materials Required.....	2
Precautions.....	3
Sample Preparation.....	3
Reagent Preparation.....	4
Assay Procedure	4
Performance Characteristics	6
Data Templates	7

Procedure Summary

- | | | | |
|---|--|--|---|
|  |  |  |  |
| 1. Add 50 μ l of standards or samples to each well in duplicate. | 2. Cover plate and incubate at room temperature (RT) for 1 hour. Do not wash or decant the plate. | 3. Add 50 μ l of Biotinylated Antibody Reagent to each well. | 4. Cover plate and incubate at RT for 1 hour. |
|  |  |  |  |
| 5. Wash plate THREE times. | 6. Add 100 μ l of prepared Streptavidin-HRP to each well. | 7. Cover plate and incubate at RT for 30 minutes. | 8. Wash Plate THREE times. |
|  |  |  |  |
| 9. Add 100 μ l of Premixed TMB Substrate Solution to each well. | 10. Develop plate in the dark at RT for 30 minutes. | 11. Stop reaction by adding 100 μ l of Stop Solution to each well. | 12. Measure absorbance on a plate reader and calculate results. |

Additional Materials Required

- Precision pipettors with disposable plastic tips to deliver 5-1,000 μ l and plastic pipettes to deliver 5-15 ml
- A glass or plastic two-liter container to prepare Wash Buffer
- A squirt wash bottle or an automated 96-well plate washer
- 1.5 ml polypropylene or polyethylene tubes to prepare standards – do not use polystyrene, polycarbonate or glass tubes and a 15 ml plastic tube to prepare Streptavidin-HRP Solution
- Disposable reagent reservoirs
- A standard ELISA reader for measuring absorbance at 450 nm and 550 nm. If a 550 nm filter is not available, the absorbance can be measured at 450 nm only. Refer to the instruction manual supplied with the instrument being used.
- Graph paper or a computerized curve-fitting statistical software package

Precautions

- All samples and reagents must be at room temperature (20-25°C) before use in the assay.
- Review all instructions carefully and verify components against the Kit Contents list (page 1) before beginning the assay.
- Do not use a water bath to thaw samples. Thaw samples at room temperature.
- When preparing standard curve and sample dilutions in culture medium, use the same medium used to culture the cells. For example, if RPMI with 10% fetal calf serum (FCS) was used to culture cells, then use RPMI with 10% FCS to dilute the standard and samples. Do NOT use RPMI without serum supplement.
- If using a multichannel pipettor, always use a new disposable reagent reservoir.
- Use new disposable pipette tips for each transfer to avoid cross-contamination.
- Use a new adhesive plate cover for each incubation step.
- Once reagents have been added to the plate, take care NOT to let plate DRY at any time during the assay.
- Avoid microbial contamination of reagents.
- Vigorous plate washing is essential.
- Avoid exposing reagents to excessive heat or light during storage and incubation.
- Do not mix reagents from different kit lots. Discard unused kit components.
- Do not use glass pipettes to measure TMB Substrate. Take care not to contaminate the solution. If the solution is blue before use, DO NOT USE IT.
- Individual components may contain antibiotics and preservatives. Wear gloves while performing the assay to avoid contact with samples and reagents. Please follow proper disposal procedures.

Additional Precautions for the 5-plate Kit

- Dispense and equilibrate to room temperature only reagent volumes required for the number of plates being used. Do not combine leftover reagents with those reserved for additional plates.
- Use only one bottle of the TMB Substrate Solution per 96-well plate. Do not combine leftover substrate with that reserved for other plates.
- Use only one vial of standard per 96-well plate.

Sample Preparation

Sample Handling

- Serum; EDTA, heparin or sodium citrate plasma; urine; or culture supernatants may be tested in this ELISA.
- 50 µl per well of serum, plasma, urine or culture supernatant are required.
- Store samples to be assayed within 24 hours at 2-8°C. For long-term storage, aliquot and freeze samples at -70°C. Avoid repeat freeze-thaw cycles when storing samples.
- Samples and standards must be assayed in duplicate each time the assay is performed.
- Gradually equilibrate samples to room temperature before beginning assay. Do not use a heated water bath to thaw or warm samples.
- Mix samples by gently inverting tubes.
- If samples are clotted, grossly hemolyzed, lipemic or microbially contaminated, or if there is any question about the integrity of a sample, make a note on the template and interpret results with caution.

Sample Dilution

- If the human IL-1 α concentration possibly exceeds the highest point of the standard curve (i.e., 400 pg/ml), prepare one or more 10-fold dilutions of the sample. When testing **culture supernatants**, prepare serial dilutions using the culture medium. When testing **serum, plasma or urine**, prepare serial dilutions using the Standard Diluent provided. A 10-fold dilution is prepared by adding 50 µl of test sample to 450 µl of appropriate diluent. Mix thoroughly between dilutions.

Reagent Preparation

For procedural differences when using partial plates, look for **(PP)** throughout these instructions.

Note: When using the 5-plate kit, only one standard per plate is supplied, therefore, partial plates cannot be used.

Wash Buffer

Note: Wash Buffer must be at room temperature before use in the assay. Do not use Wash Buffer if it becomes visibly contaminated during storage.

1. Label a clean glass or plastic two-liter container "Wash Buffer." The 30X Wash Buffer may have a cloudy appearance.
2. If using the 5-plate kit, add 30 ml Wash Buffer to 870 ml water for each plate used, otherwise, add the entire contents of the 30X Wash Buffer bottle (50 ml) to the container. Dilute buffer to a final volume of 1.5 L with ultrapure water and mix thoroughly.

(PP) When using partial plates, store the reconstituted Wash Buffer at 2-8°C.

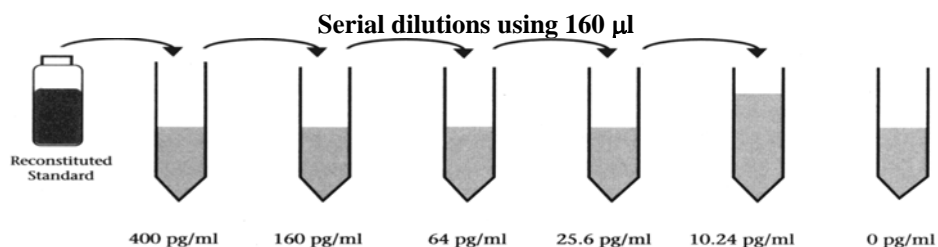
Standards

- **(PP)** Reconstitute and use one vial of the lyophilized standard per partial plate.
 - Prepare standards just before use and use within one hour of reconstitution. Do not store reconstituted standards.
1. When testing **culture supernatant samples**, reconstitute standard with ultrapure water. Reconstitution volume is stated on the standard vial label. The standard will dissolve in approximately 1 minute. Mix by gently inverting vial. Use the sample culture medium to prepare standard curve serial dilutions.

When testing **serum, plasma or urine samples**, reconstitute standard with ultrapure water. Reconstitution volume is stated on the standard vial label. The standard will dissolve in approximately 1 minute. Mix by gently inverting vial. Use the Standard Diluent provided to prepare standard curve serial dilutions.

When testing **serum, plasma or urine and cell culture supernatant samples on the same plate**, validate the medium to establish if the same standard curve can be used for the different sample types. Prepare a standard curve (including a zero/blank) using culture medium to reconstitute and dilute the standard. Use medium containing serum or other protein supplement to maximize IL-1 α stability. Perform this curve in parallel with a standard curve reconstituted in ultrapure water and diluted in the Standard Diluent provided. If OD values are within 10% of the mean for both curves, then the assay may be performed with Standard Diluent, whether testing culture supernatant, plasma, urine or serum samples.

2. Label six tubes, one for each standard curve point: 400 pg/ml, 160 pg/ml, 64 pg/ml, 25.6 pg/ml, 10.24 pg/ml, and 0 pg/ml, then prepare 1:2.5 serial dilutions for the standard curve as follows:
3. Pipette 240 μ l of appropriate diluent into each tube.
4. Pipette 160 μ l of the reconstituted standard into the first tube (i.e., 400 pg/ml) and mix.
5. Pipette 160 μ l of this dilution into the second tube (i.e., 160 pg/ml) and mix.
6. Repeat the serial dilutions (using 160 μ l) three more times to complete the standard curve points.



Assay Procedure

A. Sample and Biotinylated Antibody Reagent Incubation

- **(PP)** Determine number of strips required and leave these strips in the plate frame. Tightly seal remaining unused strips in the provided foil pouch with desiccant and store at 2-8°C. After completing assay, retain plate frame for second partial plate. When using the second partial plate, place strips securely in the plate frame.

-
- Use the Data Template provided to record locations of the zero standard (blank or negative control), human IL-1 α standards and samples. Perform five standard points and one blank in duplicate with each series of unknown samples.
 - If using a multichannel pipettor, use a new reagent reservoir to add the Biotinylated Antibody Reagent. Remove from the vial only the amount required for the number of strips being used. Take care not to touch the samples in wells with the pipette tip when adding the Biotinylated Antibody Reagent.
1. Add 50 μ l of reconstituted standards or test samples in duplicate to each well.
Note: If the IL-1 α concentration in any sample possibly exceeds the highest point on the standard curve, 400 pg/ml, see Sample Preparation – Sample Dilution section.
 2. Add 50 μ l of Standard Diluent to all wells that do not contain standards or samples.
 3. Carefully cover plate with an adhesive plate cover. Ensure all edges and strips are sealed tightly by running your thumb over edges and down each strip. Incubate for one (1) hour at room temperature, 20-25°C.
 4. Carefully remove the adhesive plate cover. Do NOT decant or wash the plate.
 5. Add 50 μ l of Biotinylated Antibody Reagent to each well containing standards or samples. Take care not to cross-contaminate the standards or samples in the plate. Mix well by gently tapping the plate several times.
 6. Carefully cover plate with an adhesive plate cover. Ensure all edges and strips are sealed tightly by running your thumb over edges and down each strip. Incubate for one (1) hour at room temperature, 20-25°C.
 7. Carefully remove adhesive plate cover. Wash plate as described in the Plate Washing section (section B).

B. Plate Washing

3. Gently squeeze the long sides of plate frame before washing to ensure all strips securely remain in the frame.
4. Empty plate contents. Use a squirt bottle to vigorously fill each well completely with Wash Buffer, then empty plate contents. Repeat procedure two additional times for a total of THREE washes. Blot plate onto paper towels or other absorbent material.

Note: For automated washing, aspirate all wells and wash THREE times with Wash Buffer, overfilling wells with Wash Buffer. Blot plate onto paper towels or other absorbent material.

C. Streptavidin-HRP Solution Preparation and Incubation

- Prepare Streptavidin-HRP Solution **immediately before use**. Do not prepare more Streptavidin-HRP Solution than required.
 - Use a 15 ml plastic tube to prepare Streptavidin-HRP Solution.
 - **Note:** If using a multichannel pipettor, **use a new reagent reservoir and pipette tips** when adding the prepared Streptavidin-HRP Solution.
1. Briefly centrifuge the Streptavidin-HRP Concentrate to force entire vial contents to the bottom.
 2. **(PP)** Use only the Streptavidin-HRP Solution amount required for the number of strips being used. For each strip, mix 2.5 μ l of Streptavidin-HRP Concentrate with 1 ml of Streptavidin-HRP Dilution Buffer. Store Streptavidin-HRP Concentrate reserved for additional strips at 2-8°C.

For one complete 96-well plate, add 30 μ l of Streptavidin-HRP Concentrate to 12 ml of Streptavidin-HRP Dilution Buffer and mix gently.

3. Add 100 μ l of prepared Streptavidin-HRP Solution to each well.
4. Carefully attach a new adhesive plate cover, ensuring all edges and strips are tightly sealed. Incubate plate for 30 minutes at room temperature, 20-25°C.
5. Carefully remove the plate cover and discard plate contents. Wash plate as described in the Plate Washing section (section B).

D. Substrate Incubation and Stop Step

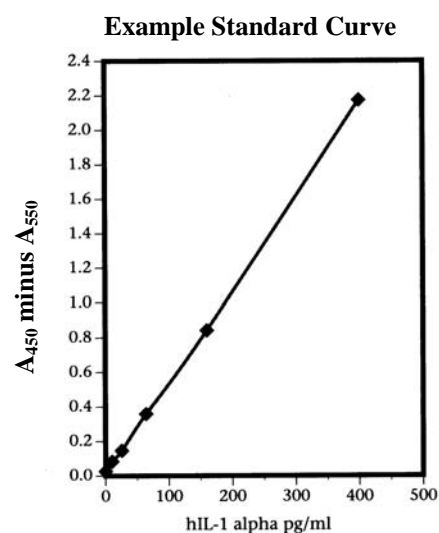
- Use new disposable reagent reservoirs when adding TMB Substrate Solution and Stop Solution.
 - Dispense from bottle ONLY amount required, 100 µl per well, for the number of wells being used. Do not use a glass pipette to measure the TMB Substrate Solution.
 - **(PP)** Do not combine leftover substrate with that reserved for the second partial plate. Take care not to contaminate remaining TMB Substrate Solution.
1. Pipette 100 µl of TMB Substrate Solution into each well.
 2. Allow enzymatic reaction to develop at room temperature in the dark for 30 minutes. Do not cover plate with aluminum foil or a plate sealer. The substrate reaction yields a blue solution that turns yellow when Stop Solution is added.
 3. After 30 minutes, stop the reaction by adding 100 µl of Stop Solution to each well.

E. Absorbance Measurement

1. Evaluate the plate within 30 minutes of stopping the reaction. Measure absorbance on an ELISA plate reader set at 450 nm and 550 nm. Subtract 550 nm values from 450 nm values to correct for optical imperfections in the microplate. If 550 nm is not available, measure absorbance at 450 nm only. Omitting the 550 nm measurement will result in higher absorbance values.

F. Calculation of Results

- Use the standard curve to determine IL-1α amount in an unknown sample. Generate the standard curve by plotting the average absorbance obtained for each standard concentration on the vertical (Y) axis vs. the corresponding human IL-1α concentration (pg/ml) on the horizontal (X) axis.
- Calculate results using graph paper or curve-fitting statistical software. Determine the IL-1α amount in each sample by interpolating from the absorbance value (Y axis) to IL-1α concentration (X axis) using the standard curve.
- If the sample was diluted, multiply the interpolated value by the dilution factor to calculate amount of human IL-1α in the sample.
- Absorbance values obtained for duplicates should be within 10% of the mean value. Carefully consider duplicate values that differ from the mean by greater than 10%.



Performance Characteristics

Sensitivity: < 2 pg/ml

The sensitivity or Lower Limit of Detection (LLD)¹ is determined by assaying replicates of zero and the standard curve. The mean signal of zero + 2 standard deviations read in dose from the standard curve is the LLD. This value is the smallest dose that is not zero with 95% confidence.

Assay Range: 10.24-400 pg/ml

Suggested standard curve points are 400, 160, 64, 25.6, 10.24 and 0 pg/ml.

Reproducibility:

Intra-assay CV: < 10%

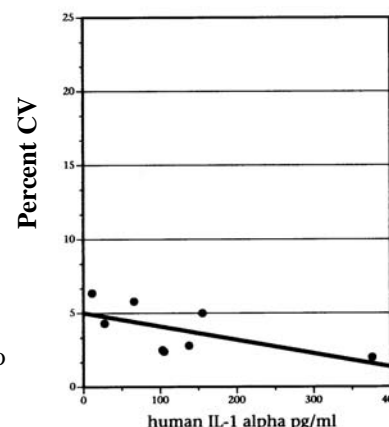
Inter-assay CV: < 10%

Specificity: This ELISA is specific for the measurement of natural and recombinant human IL-1α and does not cross-react with human IFNα, IFNγ, IL-1β, IL-2, IL-3, IL-4, IL-6, IL-7, IL-8, TNFα, or mouse IL-1α. Human IL-1ra does not interfere with this ELISA.

Calibration: Standards in this ELISA have been re-calibrated to the NIBSC reference standard lot 86/632. One (1) pg of Endogen standard = 2.5 NIBSC pg = 0.25 NIBSC units.

Precision: The inter-assay coefficient of variation is plotted against human IL-1 α concentration (pg/ml). Points represent samples evaluated in replicates of four in three different kit lots.

Precision Profile of the hIL-1 α ELISA



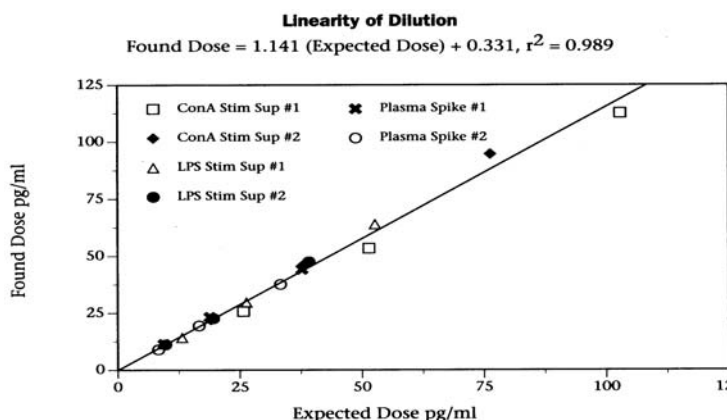
Expected Values: Serum, plasma and urine samples were tested. Ranges of normal levels of human IL-1 α are as follows:

<u>Sample Type</u>	<u>Range</u>
Serum (n=9)	0-5.4 pg/ml
Plasma (n=9)	Undetectable
Urine (n=9)	0-4.335 pg/ml

Recovery: Low, medium and high levels of recombinant human IL-1 α were spiked into serum, plasma, and urine samples from healthy individuals, as well as a control buffer. Mean recoveries are as follows:

<u>Recombinant Control</u>	<u>180 pg/ml</u>	<u>75 pg/ml</u>	<u>30 pg/ml</u>
Mean Serum Recovery	89%	89%	90%
Mean Plasma Recovery	98%	100%	99%
Mean Urine Recovery	60%	61%	69%

Linearity of Dilution: Six different positive samples were serially diluted. The dilutions are evaluated in the ELISA and “found” doses are plotted against “expected” doses.



Reference

1. *Immunoassay: A Practical Guide*, Chan and Perlstein, Eds., 1987, Academic Press: New York, p71.

This product (“Product”) is warranted to operate or perform substantially in conformance with published Product specifications in effect at the time of sale, as set forth in the Product documentation, specifications and/or accompanying package inserts (“Documentation”) and to be free from defects in material and workmanship. Unless otherwise expressly authorized in writing, Products are supplied for research use only. No claim of suitability for use in applications regulated by FDA is made. The warranty provided herein is valid only when used by properly trained individuals. Unless otherwise stated in the Documentation, this warranty is limited to one year from date of shipment when the Product is subjected to normal, proper and intended usage. This warranty does not extend to anyone other than the original purchaser of the Product (“Buyer”).

No other warranties, express or implied, are granted, including without limitation, implied warranties of merchantability, fitness for any particular purpose, or non infringement. Buyer’s exclusive remedy for non-conforming Products during the warranty period is limited to replacement of or refund for the non-conforming Product(s).

There is no obligation to replace Products as the result of (i) accident, disaster or event of force majeure, (ii) misuse, fault or negligence of or by Buyer, (iii) use of the Products in a manner for which they were not designed, or (iv) improper storage and handling of the Products.

Current versions of product instructions are available at www.thermo.com/pierce. For a faxed copy, call 800-874-3723 or contact your local distributor.

© 2009 Thermo Fisher Scientific Inc. All rights reserved. Unless otherwise indicated, all trademarks are property of Thermo Fisher Scientific Inc. and its subsidiaries. Printed in the USA.

Data Templates

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												